Poster #9

How the choice of PK model and AIF affect DCE-MRI detection of pancreatic cancer responses to stroma-directed drug

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Background & Rationale

- The dense extracellular matrix in pancreatic ductal adenocarcinoma (PDA) is a mechanism for treatment failure [1]. PEGPH20 degrades hyaluronan (HA) from ECM.
- 2. Test the utility of DCE-MRI to detect responses to stromadirected interventions
 - > DCE-metrics (K^{trans} , k_{ep} and V_p) and PK models
 - Individual arterial input function (AIF) vs. group-AIF
- 3. Corroborate imaging and immunostaining data.





H 200 μm 200 μm



IIIII $P_{out} > P_{in}$; Limited permeability
and perfusion



P_{out} < P_{in}; Permeability and perfusion increased

Group AIF (n=20)



Correlation of K^{trans} derived from SSM using Individual and Group AIF.



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Detailed MRI Methods

- > 9.4T DirectDrive[®] (Agilent) interfaced with a 12-cm gradient coil.
- ➢ RF coil: 35 mm ID x 10 cm long quadrature birdcage transceiver (M2M)
- Slice groups: one slice containing the left ventricle (LV) to obtain the arterial input function (AIF); 4-7 slices covering the tumor
- T₁₀ (T₁ before CA injection) map of tumor and blood (LV) using an ECGgated inversion recovery technique [2].
- Contrast agent (CA): MultiHance[®] diluted to 10 mM of gadolinium in saline (0.2 mL) was injected in 10 sec via syringe pump into tail vein.
- > DCE series: ECG-gated saturation recovery sequence
 - A total of 80 images were acquired continuously while CA was injected after first 10 images.
 - FOV=32 mm, matrix size = 64x64, effective TR= 2 x heart beats ≈ 200 ms, TE= 3 ms, flip angle = 90 degrees.
 - the timing of radiofrequency excitation was recorded on a microcontroller device and the record was used to correct ECG triggering errors during post processing.

Data Processing

- Individual AIF was extracted from LV heart of each mouse
- Group AIF was the average of 20 AIFs measured from 10 mice.
 - ✓ Each mouse contributed two AIFs (pre and post treatment).
 - ✓ AIFs were aligned by bolus-arrival time of CA before averaging.
- AIF, DCE series and T₁₀ maps of the tissue were fit to a pharmacokinetic model using the least squares methods
 - ✓ Tofts model [3]
 - ✓ Modified Tofts model (M-T)
 - ✓ Shutter-speed model (SSM) [4-5]
- > Pixel-wise parametric maps of K^{trans} ,
 - \textit{k}_{ep} , \textit{V}_{e} , \textit{V}_{p} and τ_{i} were obtained.

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Individual AIF





Group AIF

k_{ep} Individual Group metric: AIF AIF Tofts Yes No SSM Yes Yes

Individual AIF approach allows Tofts model to detect changes of k_{ep} induced by PEGPH20.



Tofts

SSM



% change of K ^{trans} metric:	Individual AIF	Group AIF
Tofts	No	No
SSM	Yes	Yes

Both the individual and group AIF approach only allow SSM model to detect %change of K^{trans} induced by PEGPH20.

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<i>K^{trans}</i> metric:	Individual AIF	Group AlF
Tofts	No	No
SSM	No	Yes

Only group AIF approach allows SSM model to detect *K*^{trans} change before and after PEGPH20 injection.



Immunostaining of CD31 (endothelial marker) in tumor

 V_p (fractional plasma volume) derived from fitting of M-T model using **group AIF** shows a significant increase 24 h after PEGPH20 injection. In PEGPH20-treated tumor, a trend of increase vascular lumen area is observed (arrows) compared to VEH treated tumor although a statically significance has not been reached due to small sample size.

- 1. SS model appears to be more sensitive than Tofts or M-T model for detection of treatment effect when applied with either individual- or group-AIF.
- 2. Tofts model, however, is capable of detecting detect a significant increase of k_{ep} after the treatment **only** when combined with Individual-AIF approach.
- 3. Further validation of V_p by IHC analyses is ongoing.

References

[1] S.R. Hingorani, L. Zheng, A.J. Bullock, T.E. Seery, W.P. Harris, D.S. Sigal, F. Braiteh,
P.S. Ritch, M.M. Zalupski, N. Bahary, P.E. Oberstein, A. Wang-Gillam, W. Wu, D.
Chondros, P. Jiang, S. Khelifa, J. Pu, C. Aldrich, A.E. Hendifar, J Clin Oncol, 36 (2018)
359-366.

[2] J. Cao, S. Pickup, C. Clendenin, B. Blouw, H. Choi, D. Kang, M. Rosen, P.J. O'Dwyer, R. Zhou, Clin Cancer Res, 25 (2019) 2314-2322.

[3] P.S. Tofts, J Magn Reson Imaging, 7 (1997).

[4] R. Zhou, S. Pickup, T.E. Yankeelov, C.S. Springer, Jr., J.D. Glickson, Magn Reson Med, 52 (2004) 248-257.

[5] X. Li, W.D. Rooney, C.S. Springer, Jr., Magn Reson Med, 54 (2005) 1351-1359